INVESTIGATION OF THE FREE FORMALDEHYDE CONTENT
WHEN POLIOMYELITIS VIRUS IS INACTIVATED
WITH FORMALIN

From the Biochemical Laboratory (Head — Candidate Biol. Sci. N. V. Kholchev) of the Moscow Research Institute of Antipoliomyelitis Preparations (Scientific Director — Prof. V. D. Solov'ev)

(Received October 16, 1958, Presented by Active Member AMN SSSR A, E, Braunshtein)

The considerable interest in the inactivation of viruses with formalin is due primarily to the preparation of antipoliomyelitis vaccine. Of the more important conditions of the inactivation process [4] which have not been considered quantitatively, there has remained the quantity of formaldehyde reacting with the virus. It is known that not all the formaldehyde added reacts with the virus, but that part of it is fixed by other components of the virus-containing fluid—amino acids, peptides and proteins—which pass into it from the synthetic medium 199 (Parker) and accumulate in the culture tissue in the process of proliferation. This bond is largely reversible.

In order to judge the quantity of formaldehyde capable of causing inactivation without essential depression of antigenicity, two values are therefore required—the total and the free formaldehyde.

Until recently, however, there has been no sufficiently reliable quantitative method of estimation of free formaldehyde [2, 8]. The only known indication was that of Taylor and Moloney [7] that, from the results of one approximate diffusion experiment, the content of free formaldehyde in monovalent poliomyelitis vaccine was about 90 %.

The method which we have devised for the determination of free formaldehyde in the presence of labile amino acid-formaldehyde and protein-formaldehyde compounds has enabled us to obtain the necessary values for the solution of this problem.

EXPERIMENTAL METHOD

We investigated different, independently prepared batches of virus fluid, obtained from a culture of kidney tissue in synthetic medium 199 and inactivated in accordance with Salk's [4, 5] recommendations by the addition of 1:4000 formalin, giving a final formaldehyde concentration of 0.1 g/l.

In each case a sample for testing was taken on the 12th day of storage of the inactivated fluid at a temperature of 37°, i.e., in the phase of completion of the reaction between formaldehyde and the components of the virus fluid. The amino-nitrogen was estimated before its partial combination with formaldehyde, in the original virus fluid without formalin. The method of estimation of free formaldehyde has been described previously by us [1], and we shall merely give here a brief account of the principle of the analysis.

The investigation was carried out in a modified Conway's dish, in which the base of the inner chamber was slightly raised so that its depth was 1.5-2.0 mm. The outer chamber was filled with the test solution. A small volume of water (0.3 ml), covering the base of the inner chamber in a thin film, absorbed the gas evolved from this solution. In a state of equilibrium (after 42 hours) a concentration of free formaldehyde was produced in the absorbing liquid in which the presence of labile formaldehyde compounds did not interfere with the subsequent estimation of the formaldehyde by a colorimetric method [6].

TABLE 1
Free Formaldehyde Content in Inactivated Virus Fluid

Serial No.	No. of virus	Free form= aldehyde (in g/1)	Serial No.	No. of virus	Free form- aldehyde (in g/1)
1 2 3 4 5 6 7 8 9 10 11 12 13	30 31 32 33 35 43 44 44 48 49 50 51 52 53	0,071 0,081 0,036 0,081 0,075 0,089 0,086 0,080 0,085 0,080 0,085 0,076 0,079	15 16 17 18 19 20 21 22 23 24 25 26 27 28	55 56 57 58 59 60 62 66 67 68 69 70 71	0,080 0,076 0,085 0,075 0,086 0,079 0,080 0,084 0,081 0,072 0,078 0,078 0,078

Mean - $0.080 \pm 0.009 \text{ g/1}$, $\delta = 0.0044$, medium 199+ formalin 1: 4000 - 0.086 g/1.

TABLE 2

Total Nitrogen Content of Inactivated Virus Fluid

Serial No.	No. of virus fluid	Total nitrogen (in mg/1)	
	1	1	
1	31	147	
2	35	147	
2 3 4 5	43	159	
4	448	147	
5	44Zh	151	
6	48	140	
7	49	149	
8	50	140	
9	51	152	
10	52	150	
11	53	151	
12	55	135	
13	56	143	
14	57	140	
Mean		147±12	

Medium 199	139
N-40 Who Starre of 150 month	-6

 $\sigma = 6, 1$

Note: The figure of 156 mg/l of total nitrogen in the virus fluid before addition of solutions of formaldehyde and acetic acid corresponds by calculation to the mean total nitrogen content in the monovalent vaccines – 147 mg/l.

TABLE 3

Amino - Nitrogen Content of the Virus
Fluid

Serial No.	No. of virus fluid	Amino- nitrogen (in mg/1)	
1 2 3 4 5 6 7 8 9 10 11 12 13	32 33 43 44 S 44 Zh 60 62 65 69 70 71 72 73 75	126 117 124 127 121 122 115 126 122 120 120 120 122 121	
Mean		121 <u>+</u> 6 σ==3,4	

Medium 199 116

Note: Amino-nitrogen content of medium 199, by calculation 112 mg/1.

The total formaldehyde was estimated by the Haiberger - Nitschmann [3] method, the total nitrogen by Conway's method and the amino-nitrogen by the ninhydrin method.

EXPERIMENTAL RESULTS

It will be seen from the figures in Table 1 that the free formaldehyde content in the different batches of inactivated virus fluid was on the average 0.08 g/l. It is interesting that the medium 199 itself combined with almost tha same amount of formaldehyde, leaving 0.086 g/l uncombined.

It may be accepted that the formaldehyde in the virus fluid is combined mainly with the amino acids of medium 199, which do not undergo any essential quantitative changes in the process of cultivation of the virus (see also Tables 2 and 3).

Estimation of the total formaldehyde content of 5 batches immediately after addition of formalin showed that its value averaged 0.098 g/1 (0.096-0.101 g/1), and in 20 batches at the end of the process of inactivation its value was 0.097 g/1 (0.091-0.107 g/1).

The free formaldehyde thus accounted for 82.5% of the total. The considerable surplus of free formaldehyde reduced to a minimum the effect of small variations arising from treatment of the virus fluid, and ensured correct reproduction of its concentration.

In view of the practical importance of reproduction of the concentration of free formaldehyde, it appeared desirable to verify this conclusion by comparison with those chemical indices which reflect the concentration of those substances in the virus fluid that react with formaldehyde. The total concentration of these substances was judged by the total nitrogen present in the monovalent vaccines, and of the individual functional groups we investigated the most important — the amino group.

It may be seen from the figures in Tables 2 and 3 that the concentrations of the substances fixing formaldehyde were equally stable as the free formaldehyde. Hence it follows, in particular, that the practical recommendations directed towards the compensation of the supposed instability of the free formaldehyde concentration, and the resulting absence of standardization of the process of inactivation, for example the addition of glycine [2, 8] or further addition of formalin, have no sound basis.

We were thus able to establish that during the inactivation of the virus of poliomyelitis with formalin in accordance with the accepted recommendations [4,5] for preparation of the vaccine, the free formaldehyde content in the inactivated virus fluid had an average value of 0.08 g/1, and this concentration was readily reproduced in practice.

SUMMARY

The quantitative diffusion method of determination of free formaldehyde has established that, inactivating poliomyelitis virus by Salk's method, the average free formaldehyde level at the end of the process was 0.08 g/l.

Formaldehyde in the virus fluid is combined mainly with amino acids passing into the fluid from the 199 medium. The free formaldehyde concentration is an easily controlled condition of inactivation.

This is confirmed by the stability of the chemical indices, the level of total nitrogen and amino nitrogen, reflecting the level of the substances interacting with formaldehyde in the virus fluid.

LITERATURE CITED

- [1] M. Ya. Fel'dman, Biokhimiya, 23, No. 6, 917 (1958).*
- [2] S. Gard, The Nature of Viruses, pp. 135-157, Moscow, 1958 [Russian translation].
- [3] H. Nitschmann, H. Hadorn, Helv. chem. acta, c 24, p. 237-242 (1941), cited in: D. F. Walker Formaldehyde, Moscow, 1957 [Russian translation].
 - [4] J. E. Salk, U. Krech, J. S. Younger et al., Am. J. Pub. Health v. 44, p. 563-570 (1954).
 - [5] J. E. Salk, Am. J. Pub. Health v. 46, p. 1-14 (1956).

^{*} Original Russian pagination. See C.B. Translation.

- [6] M. Tanenbaum, C. E. Bricker, Analyt. Chem. v. 23, p. 354-257 (1951).
- [7] E. M. Taylor, P. J. Moloney, J. Am. Pharm. A. Sc. Ed. v. 46, p. 299-301 (1957).
- [8] T. Wesslen, E. Lycke, S. Gard et al., Arch. ges. Virusforsch. Bd. 7, S. 125-135 (1957).